

1 Immunophenotyping of Acute Lymphoblastic Leukemia in
2 Sudanese Children's Versus Egyptian Children's using
3 Flowcyometry

4 Dr. Amged Hussein Abdelrahman

5 *Received: 6 February 2021 Accepted: 28 February 2021 Published: 15 March 2021*

6

7 **Abstract**

8 Background: Acute lymphoblastic leukemia (ALL) develops rapidly, creating immature white
9 blood cells (WBCs) called lymphoblasts. This type of hematological malignancy is cancer
10 involving blood and bone marrow (BM). It can affect both B and T cell lineage. Here we
11 studied T B markers in 100 Sudanese ALL patients attending the Radiation Isotopes Center
12 Khartoum versus 180 Egyptian ALL patients attending the Alksr Aani, oncology center, Cairo
13 between October 2009 and August 2014. The study aimed to detect the frequency of B and T
14 lineages subclass in ALL among the Sudanese versus Egyptian population in correlation with
15 their clinical symptoms, hematological parameters, gender and, age.

16

17 **Index terms**— sudanese, egyptian children, acute lymphoblastic leukemia, flowcytometry.

18 **1 Immunophenotyping of Acute Lymphoblastic**

19 Leukemia in Sudanese Children's Versus Egyptian Children's using Flowcyometry Dr. Abdelgadir Ahmed
20 Abdelgadir ?, Dr. Khalid Omer Abdallah Abosalif ?, Dr. Abozer Ahmed Alderdery ? & Dr. Amged Hussein
21 Abdelrahman ?

22 The study aimed to detect the frequency of B and T lineages subclass in ALL among the Sudanese versus
23 Egyptian population in correlation with their clinical symptoms, hematological parameters, gender and, age.

24 Materials and methods: A questionnaire was used to collect geographical information and, clinical data. Initially,
25 full blood count (FBC), peripheral blood morphology and BM examination was done to diagnose patients. FBC
26 was done by a haematology analyzer and Leishman stain was used for the cell morphology. Immunophenotype CD
27 markers, TdT, CD3cyt, CD3 surf, CD5, CD7, CD2, CD10, CD19, CD20, Ig, sIg, and CD2 antigens were looked
28 for to determine the B and T subclasses of ALL using flow cytometry. Mononuclear cells were prepared for flow
29 cytometry and labeled with fluorescence conjugated antibodies. Analyzing gated populations with EPICS XL
30 using three-color protocols, fluorescence intensity and peak width were calculated for each antigen. TdT, CD10
31 and, CD19 were positive in all subclasses of B lineage ALL. ((sIg and Ig-cyto were detected in B ALL and Pre B
32 ALL were detected the former was sIg. In contrast, sIg and Ig-cyto markers were not detected in cases with Pro
33 B ALL. Panel T CDs were detected in thymocyte and post thymocyte ALL, as the former was positive CD3 (cyto)
34 and the latter was positive CD3 (surf). TdT, CD2 and CD5 immunophenotypes were positive in all subclasses of
35 T-ALL)). Pro B ALL was diagnosed in 24% (n=43) and 23% (n=23) of cases, 51% (n=91) and 29% (n=29)
36 were Pre B ALL, 9% (n=16) and 29% (n=29) BALL, 4% (n=8) and 11% (n=11) thymocyte ALL and, post
37 thymocyte ALL was 12% (n=22) and 9% (n=9) among Egyptian and, Sudanese patients respectively.

38 Result: Patients were classified into three categories based on their ages, (1-4 Yrs), (5-8 Yrs) and
39 (9)(10)(11)(12). The frequency of ALL in the first group was significantly higher in Egyptian than in Sudanese
40 children ($p<0.05$), while in the last group (9-12 yrs), it was significantly higher in Sudanese children ($p<0.05$).
41 Pre-B ALL was significantly higher in Egyptian than in Sudanese ($p<0.05$), particularly in those less than 8Yrs,
42 whereas BALL was significantly higher in Sudanese in comparison with Egyptian ($p<0.05$). With regards to
43 prevalence of T lineage ALL (thymocyte and post thymocyte) and, cases of B lineage ALL cases (Pro, Pre and
44 B ALL), there was no significant variation ($p>0.05$) between the two ethnic groups Sudanese and Egyptian.

5 MATERIALS AND METHODS

45 Their significance is represented in parenthesis. You mentioned a significant difference. Regarding hematological
46 parameters, the means of Hb concentration, TWBC, platelets and, lymphoblasts for patients with ALL were
47 9 and 8 g/dl, 37X 10³/cm and 32X10³, 64X10³/cm and 65X10³ /cm and, 75%and 72% in the Sudanese and,
48 Egyptian respectively. The presence or absence of lymph node were stratified this way, pro B ALL, pre B ALL,
49 B ALL, thymocyte ALL and post thymocyte. Their presence were (12%) , (13%), (14%), (5%) and (8%) In the
50 Sudanese patients while in the Egyptians it was (19.5%), (44%), (7%),(9.5%) and (4.5%). Flow cytometry has a
51 distinctive role in the diagnosis and differentiation of ALL. Diverse flow cytometric parameter use helps minimize
52 marker numbers leading to reduced cost without reduced accuracy.

53 2 Conclusion:

54 Age ranging from one year to 12 years with a mean of 6.5 years. The male to female ratio was 1:37 High age group
55 in Egyptian ethnic group was 1-4 Yrs while in the Sudanese ethnic group, the higher age group was 9-12 Yrs. In
56 this study, B lineage origin is the most common type than T lineage origin in two ethnic groups: the T lineage
57 had a better prognosis than B lineage. In this study, also thymocyte ALL with cytoplasmic CD3 in the pediatric
58 group below two years showed with the high total leucocytic count. Flow cytometry has a distinctive role in
59 the diagnosis and differentiation of ALL.using of certain flow cytometric parameters can helps in minimization
60 of cost without reduced accuracy. There is significant variations in ALL subclassification between Sudanese and
61 Egyptian Patients that may be due to genetic background.

62 3 Introduction

63 ontemporary research on childhood acute lymphoblastic leukemia (ALL) has focused on the identification
64 of biological and clinical prognostic markers to generate better risk-adapted treatment strategies (1) The
65 identification of several cluster C differentiation markers and early diagnosis allowed the definition of patient
66 subsets with distinct prognostic features (1) Nevertheless, treatment itself remains one of the strongest prognostic
67 factors, as has been shown in several well-designed large clinical trials (2) . Cytometry has evolved from a
68 promising new technology to an indispensable tool in the diagnosis of hematologic malignancies. Many new
69 antibodies, improved gating strategies, and routine use of multiparameter techniques have dramatically improved
70 the diagnostic utility of flow cytometry. This review will focus on the use of flow cytometry in the routine
71 clinicopathologic approach to the diagnosing of leukemias and lymphomas, emphasizing the relevant literature of
72 the past ten years. Some of the recent advances in flow cytometric monitoring of disease and treatment are shown
73 in the last section. We will review the use of flow cytometry in the diagnosis of major disorders highlighting
74 the prognostically important subgroups defined either morphologically or genetically. The discussion will focus
75 not only on the use of flow cytometry in the differential diagnosis of a particular disorder, but also correlate
76 immunophenotypic, with clinical features, HB, TLC. Platelets and accumulation of blast cells. In the delineation
77 of biologically important subgroups. We intent that this review supports a combined modality approach to
78 the daily practice of hematology-oncology and hematopathology. A working knowledge of the basics of flow
79 cytometry is assumed; thus, technical aspects of instrumentation, normal distribution of surface antigens, and
80 methodologies. leukemia is a group of neoplastic diseases of blood-forming cells of the bone marrow, which result
81 in the proliferation and accumulation of immature and generally defective blood cells in both the blood-stream and
82 the bone marrow (3) . This may result in anemia, thrombocytopenia, and granulocytopenia and, infiltration of
83 other sites such ,as lymph nodes, kidney, spleen, testes, and the central nervous system (CNS). The cells involved
84 are usually leukocytes, but several different forms of the disease may be manifested; according to which leukocyte
85 cell line is involve, the leukemia are universally fatal if untreated, generally due to complications resulting from
86 the leukemic infiltration of the bone marrow and replacement of normal hematopoietic precursor cells. These
87 fatal complications are usually hemorrhage and infection (3) . Leukemia is the most common childhood cancer,
88 accounting for one-third of malignancies in children under 15 years of age in Europe and North America. The
89 annual incidence in the United Kingdom is 3 0-40 cases per million children. About 80% are acute lymphoblastic
90 leukemia (ALL) and 18% acute myeloid leukemia (AML) (4) . Acute lymphoblastic leukemia, with a sex ratio of
91 1 .2 males to females, shows a peak incidence in childhood between the ages of 3 and 5years and is due to cases
92 of early B cell ALL. T-ALL, with a male predominance, is more common in older children ('Stiller and Draper
93 1998). Both incidence and mortality are slightly higher in males (5) . A peak of age occurs between 2-4 years
94 (Margolin and Poplack 1997). in Egypt, acute leukemia is the most common pediatric malignancy accounting
95 for about 40% of childhood cancer, with ALL counting for 70% of the cases (5) . the peak incidence is between
96 3-7 years (6) .

97 4 II.

98 5 Materials and Methods

99 Study Participants; This study included 180 Egyptian children and 100 Sudanese children newly diagnosed with
100 acute lymphoblastic leukemia, 158 males and 122 females. Their age ranged from one year to 12years.

101 6 Sample preparation and methods

102 For each patient, the following samples were collect: 4ml EDTA blood sample was collected under complete
103 aseptic conditions for CBC and BM aspirate for morphological examination and immunophenotyping on EDTA
104 containers. BM samples were processed within a few hours because CD antibodies monoclonal have short
105 stabilities even if stored in the refrigerator. In most cases, samples were processed within 6 hours of collection.

106 Methods; Complete blood count Was done by using Sysmex KX-21N, Kope, Japan. Principle of smear
107 preparation A small drop of blood is place near the froste end of a clean glass slide. A second slide is used as a
108 spreader. The blood is streaked in a thin-film over the slide. The slide is allow to air-dry and is then stained. (7)
109 . Staining of thin film Leishman stain was used in staining of all blood smears and bone marrow in this study.,
110 Flowcytometry, The EDTA anticoagulated BM sampled were diluted 1.3 with PBS then the cells were stained by
111 adirect immuno-florescent technique by addition of monoclonal antibodies (microwell Test Kit contains) which
112 contains T lineage panel monoclonal antibodies (CD2, CD5, CD3 cyt, CD7 and, CD3surf) and B lineage panel
113 monoclonal antibodies (CD10, CD9, CD20, CD19 Ig cyt and, Ig surf.) and Leukocyte marker (TdT). Sample
114 staining be carried out as soon as possible after the nucleated cell suspension has been prepared. Delaying
115 this step will only reduce viability and induce cell clumping, especially if the tubes holding the cell suspensions
116 will be stored in an upright position (7). Direct immune fluorescence double staining Tubes was labeled with
117 the name of patients, type of the specimen, laboratory number and combination of fluorochrome. 100 μ l of
118 specimen (whole peripheral blood or bone marrow) was placed in a labeled tube., 2ml of phosphate buffered
119 saline PBS, (PH 7.3) was added containing 0.02% sodium azide 0.02% Bovine albumin., Tube was centrifuged
120 at 2000 rpm for 5 minutes and, the supernatant were removed., The cells was resuspended in 02-0.5 ml of fluid
121 sheath solution (e.g . isotonic). Then the tube was readied on a flow cytometer instrument . (8) . Detection of
122 surface Immunoglobulin, Surface 1g heavy and light chain can be detected using Double or triple immune staining
123 the object in to demonstrate clonality of a B cell population. Double staining was done by Combination of an
124 FTC labeled B cell, marker e.g, CD19 and, PE Labeled anti-light chain. The tube was label with the name of
125 the patient, type of specimen, laboratory number and MCAP. 100 μ L of the specimen (whole peripheral blood or
126 bone marrow) was pipette. 2 ml of lysine solution was added, then incubate for 10 minutes at room temperature.
127 The tube was washed twice in PBS aside from BSA. An appropriate Volume of MC Ab was add according to
128 the manufactures recommendation. The tube was re incubated for 10 minutes at room temperature. 2ml of PBS
129 aside BSA or Henks Solution was added. tube was centrifuged at 2000 rpm and, the supernatant was remove.
130 The cell was re suspended in 0.2-0.5 ml of sheath fluid (isotonic) and inserted on a flow cytometer. Detection
131 of intracellular antigens, There are several commercially available kits containing solutions to fix and stabilize
132 cells in to detect cytoplasmic and or nuclear antigens, overall, these reagents have little or no effect on the light
133 scatter pattern. Also, their reliability and consistency for detecting particular nuclear and cytoplasmic antigens
134 may vary. The kits contain two solution. A is a fixing solution. B is a stabilizing agent. The tube was label
135 with the name of the patient, type of specimen laboratory number and, the MC Ab. 100 μ L of the specimen
136 was pipette into the tube. 100 μ L of Solution A (fixative) was added and incubated at room temperature for 10
137 minutes. Tube was washed twice in PBS and BSA by centrifuging for 5 minutes at 2000 pm. 100 μ L of solution
138 B (stabilizing) and the appropriate amount of fluoromecen conjugated Mc Ab were added. Cell was incubated
139 at room temperature for 15 minutes .The cell was washed twice in PBs azide BSA by Centrifuging for 5 minutes
140 at 2000 rpm. The cell was resuspended in 0.2-0.5 ml of sheath fluid solution (isotonic) and the tube was inserted
141 on flow cytometer.

142 Statistical Analysis: Statistical assessment was carried out with statistical package for social sciences (SPSS)
143 version 17.0 for windows statistical software.

144 7 III.

145 8 Results

146 9 Age comparison within the study population

147 In this study, Sudanese and, Egyptian patients with ALL were classified into three categories based on their ages,
148 (1 to 4) Yrs, (5 to 8) Yrs and, (9 to 12) Yrs. Two hundred eighty study participants with ALL, of whom 36%
149 (n=100) were Sudanese and, 64% (n=180) were Egyptian. The frequency of (1-4 yrs) age group was significantly
150 higher in Egyptian than in Sudanese (p<0.05). In contrast, the prevalence of (9-12 yrs) age group was significantly
151 higher in Sudanese compared to Egyptian (p<0.05). The Pre-BALL was significantly higher in Egyptian than
152 in Sudanese (p<0.05), particularly in those less than 8Yrs, whereas BALL was significantly higher in Sudanese
153 in comparison with Egyptian (p<0.05). With regards to the prevalence of T lineage ALL (thymocyte and post
154 thymocyte) and cases of B lineage ALL cases (Pro, Pre and BALL), there was no significant variation (p>0.05)
155 between the two ethnic groups. Sudanese and, Egyptian. Their significance is represente in parenthesis . Table
156 (1).

157 Gender comparison within the study population In this study the majority of children with ALL were males
158 n=170(61%), 66 (66%) were Sudanese and 104(58%) were Egyptian compared to females no=80(39%), where
159 34 (34%) were Sudanese and 76 (42%) were Egyptian. The Pre-B ALL cases among Egyptian females (patients)
160 were significantly higher than in Sudanese females (patients) (p<0.05), in contraste the prevalence of B-ALL was

13 DISCUSSION

161 significantly higher in Sudanese compared to Egyptian (p<0.01). Their significance is represented in parenthesis.
162 Table (2).

163 10 Subclasses of B and T acute lymphoblastic leukemia and 164 their frequencies within the study population

165 The frequency of B lineage ALL subclasses for the two ethnic groups was 82% out of all studied samples, which
166 was approximately five times higher than those with T lineage ALL subclasses. Pro-BALL was diagnosed in 24%
167 (n=43) and 23% (n=23) of cases, 51% (n=91) and 29 % (n=29) were Pre B ALL, 9% (n=16) and 29% (n=29)
168 BALL, 4% (n=8) and 11%. Table (3).

169 11 Immunological findings in the study population

170 TdT, CD3cyt, CD3 surf, CD5, CD7, CD2, CD10, CD19, CD20, Ig cyt and, sIg antigens were investigate in the
171 differential diagnosis of B-ALL and T-ALL. TdT, CD10 and, CD19 were positive in all subclasses of B lineage
172 ALL. sIg and Ig-cyto are crucial in identifying B ALL and, Pre B ALL as the former is sIg positive and the latter
173 is positive for Ig-cyto. In contrast, sIg and Ig-cyto-markers were not detect in cases with Pro B ALL. Panel T
174 CDs were important in identifying thymocyte and post thymocyte ALL, as the former is positive for CD3 (cyto)
175 and the latter is positive for CD3 (suf). TdT, CD2 and, CD5 immunophenotypes were positive in all subclasses
176 of T-ALL. Table (4).

177 Hematological parameters of study population; Four hematological parameters were estimated; Hb, blast cells,
178 TLC and, Plts. This study shows comparative statistics between B and T lineage ALL patients of Sudanese and
179 Egyptian ethnicities. The results were not within the published normal range among study population, as Hb, PLT
180 were low in both ethnic groups and TLC, blast cells were high among them. The difference in these parameters
181 was not significant in Sudanese versus Egyptian patients p = >0.05. Table (5).

182 French American British classification: Patients were classified based on the FAB classification: 27% and
183 17.2% as L1; 62% and 77% as L2 and 11% and 5% as L3 in Sudanese and ,Egyptian respectively. Of Egyptian
184 patients, L2 was highly significant (p < 0.05) compared to other classes, L1, of Sudanese patients, L2 was highly
185 significant (p < 0.05) compared to other classes while L3 no significant variation in Sudanese and Egyptian.
186 Table (6).

187 12 Lymph node in study population:

188 To detect either presence or absence of lymph node. Stratified this way, pro B ALL, pre B ALL, B ALL,
189 thymocyte ALL and. post thymocyte. Their presence in Sudanese were (??2), (13%), (14%) ,13 (13%), (5%)
190 and (8%) while in Egyptian were (19.5%), (44%) ,(7%) ,(9.5%) and (4.5%). table (7).

191 Bleeding; To detect either presence or absence of lymph node and, to bleed for participants. Its presence was
192 stratified this way, pro B ALL, pre B ALL, B ALL, thymocyte ALL and post thymocyte. table (8).

193 IV.

194 13 Discussion

195 A cross-sectional case-control study was carried out at the Radiation & Isotopes Center, Khartoum and the Alksr
196 Aani, oncology center, Cairo to compare the prevalence of ALL immuno phenotypes amongst the Sudanese and, the
197 Egyptian population using flow-cytometry. Blood cell morphology, cytochemistry stains, cytogenetic studies and
198 immunophenotyping are basic methods for ALL diagnosis (1, 2). These techniques have been used extensively
199 over several years in different parts of the world and, a wide distribution of hematological malignancies found
200 for different regions (3)(4)(5). The data report here are based on flow-cytometry results from blood samples
201 taken from diagnosed Sudanese and Egyptian ALL patients (figure1). Flow cytometry was used to evaluate the
202 different types of ALL, B and, T lineages in the peripheral venous blood and, the detection of their subclasses,
203 based on the Cell 'Cluster of differentiation (CD) markers (6,7). Here, TdT, CD3cyt, CD3 surf, CD5, CD7, CD
204 2, CD10, CD19, CD20, Ig cyt and, sIg antigens were investigated in the differential diagnosis of B-ALL and
205 T-ALL lineages and their immunophenotypes, as discussed in Table (4). CD markers are a helpful method to
206 recognize a specific cell population, however; they might be express on more than one cell type (8,9). This was
207 also found here among study patients with T lineage, as the CD7+ was detected in 47% and 51% of Sudanese and,
208 Egyptian, respectively (See Figure 3). Thus flow-cytometry methods have been develop for immunophenotyping
209 cells with two or more antibodies simultaneously to diagnose subpopulations of ALL effectively (10,11). Of these
210 markers, sIg, CD10, CD19, CD20 and Ig cyto were used in this study is the differential diagnosis of B lineage ALL
211 immunophenotypes. Similar markers have been used in reported studies in the literature review (7,12,13). The
212 immunophenotypes of B lineage ALL can be differentiate by analyzing the results for just five CD markers, sIg,
213 Ig-cyto, CD10, CD19 and, TdT. As the TdT is positive in all immunophenotypes of ALL, the current study used
214 the first four CD markers in differentiating subclasses of B lineage ALL as follows: CD10+ and CD19+ were found
215 in all subclasses of B lineage-ALL, but they were not found in all immunophenotypes of T lineage ALL; sIg+ was
216 found only in B-ALL and Ig-cyto+ was found only in Pre B ALL. In contrast, sIg+ and Ig-cyto+ markers were not
217 found in cases with Pro B ALL, as discuss in Table ??4. This classification is important for the identification of

the outcome of the ALL immunophenotypes (14,15) (18,19). Regarding panel T lineage CD markers in the current study, CD2+ and CD5+ were found in both thymocyte and post thymocyte ALL sub populations but CD3 (cyto) + was found to be positive only for thymocyte ALL and CD3+ (suf) was positive only for post thymocyte ALL (See Table ??). In contrast, this finding was similar to the study by Yoneda, N. and co-workers concerning the presence of the CD2+ and CD5+ in T lineages ALL subpopulations (7). Of the study samples with T ALL, CD7+ was found in approximately half cases of Sudanese and Egyptian. This marker was found in all cases with bad outcomes for T ALL subpopulations, thymocyte and, post thymocyte (13,20). T lineage ALL (thymocyte &post thymocyte) also has considered with bad prognosis when is associated with t(11;14) & t(10;14) respectively (19,21). All cases had TdT at the time of initial diagnoses, but other CDs marker significantly increased during the staging of the diseases, such as Ig and CD7. Immunophenotypes also appear to affect the prognosis of ALL (figure ??). It is hoped this study may act as a pilot study to highlight the need to implement a flow cytometry for ALL and other hematological malignancies in Sudan. In the current study, B lineage ALL subclasses were detected more than T lineage ALL, but the former was a little bit more in Egyptian compared to Sudanese, 84% and 80%, respectively. Controversially, T lineage ALL subclasses were detected higher among Sudanese (20%) than Egyptian ethnicities (16%). (See Figures 2 and 3). This finding is in disagreement with other ethnicities, as in patients with ALL from Brazil and Japan. In Brazil, the B lineage ALL was detected in lower frequency 56.7% and the T lineage ALL in higher frequency 43.3% (22), while in Japan, the T-lineage ALL accounted for lower frequency (13%) and B-lineage ALL accounted for higher frequency (87%)(37) compared to the current study groups. The higher percentage of B lineage ALL in this study might explain that the outcome of ALL might worsen in Egyptian and Sudanese; as it was reported to have a significantly poor event; low survival compared with patients with B lineage ALL (20). Accurate immunophenotyping of ALL is essential to evaluate the value of treatment in early diagnosis and to individualize treatment protocols, as described in the literature review (22), the frequency of ALL subclasses constitutes the theme of this study, using CDs markers (See Table ??). Pro-B-cell ALL in all age groups is associated with an unfavorable prognosis Patients with pro-B cell phenotype had a more favorable prognosis compared to those patients with pre-B cell phenotype, based on their clinical symptoms (36). Another study reported that there was a significant correlation between immunophenotyping at diagnosis and higher complete remission rate and longer survival (24). Of Egyptian patients with B-lineage ALL, the commonest immunophenotype was pre-BALL compared to other Blineage ALL. In contrast, out of Sudanese patients with B-lineage ALL, the prevalence of B-lineage ALL immunophenotypes was approximately similar with some varying degrees (See Table ??). Accordingly, most Egyptian cases with B-lineage ALL may have bad prognosis, as most of their cases were pre B ALL as opposite of Sudanese cases with B-lineage ALL. Therefore, from the above-mentioned studies and the current findings, the immunophenotyping at diagnosis may predict the a good outcome. The quantity of CD marker was used to evaluate the outcome and staging of ALL, it found that cases with positive CD10 had a good prognosis (25). In this study, the CD10 was detect positive qualitatively in all cases with B lineage ALL. Hence, further researchers might be useful to detect CD10 quantitatively in these ethnicities. The flow cytometry technique was preferred over alternative techniques in this study because of its accuracy and reliability. It is currently reported as one of the most reliable methods for hematological malignancies; it also has the advantage of diagnosing patients with ALL and other hematological malignancies (15,24). Several antibodies must be used together to evaluate unique cell markers (35), Therefore eleven CD markers were use in the current study to immunophenotype ALL, B and T populations and subpopulations, as demonstrated in Tables ?? and 4. Although flowcytometry is currently the best available method for ALL immunophenotypes determination, it is expensive of equipment purchase and maintenance. The study is also concern with the high prevalence of ALL in Sudan, as reported previously (25,26). The data collected was also intended to instigate the relationship between immunophenotypes of ALL between Sudanese and, Egyptian patients (See Table ??). Interviews and questionnaires were designed to collect demographic: age, sex, ethnicity (tribe), family history and clinical data (See Appendix-1). This information was taken from the patients' parents and, information on clinical symptoms, family history and geographical data. ALL is very prevalent in Sudan and Egypt with, high mortality and morbidity rates (25)(26)(27)(28). It is worth searching of ALL in Sudan, as it is a fertile and , virgin area due to a lack of researches and the last international published study was done approximately three decades ago by Ahmed and, co workers (25). Thus, the current research intended to identify the frequency of B and T lineages ALL and their subpopulations in Sudanese children with ALL. Of Sudanese cases here, 17% had T lineage ALL and 81.5% had B lineage ALL (See Table ??). In contrast, this finding was similar to the study by A. Redalliland coworkers, concerning the widespread presence of the B and T lineages ALL in Italy, the United States (US), Switzerland, and Costa Rica, where ALL was report with the highest incidence (29). Furthermore, in the current study, the prevalence of ALL subclasses among Sudanese was slightly different from the Egyptian findings, 20% were T ALL and 80% had B ALL (See Table ??). The former was marginally higher in Egyptian than in Sudanese, whereas the latter was vice versa. Generally, in both ethnic groups, Sudanese and, Egyptian, the frequency of ALL was nearly similar with some varying degrees compared to the previously reported studies (29,30). In general, the prevalence of B lineage ALL was higher than T lineage ALL phenotypes (31) (32), as a study found that its frequency was 76.8% pre B and 6% as pro B and 2.3% as T ALL (19) what about the rest ?? and , another study was detected the T ALL only 1.3% in newly diagnosed ALL patients which was slightly lower than the previous study. In the current study, T ALL was 20% in Sudanese and 16% in Egyptian, which was higher compared to the above two studies (12) (15) and

281 a little bit than in a study reported by Pieter Van and coworkers (31). BALL was 80 % in Sudanese and 84%
282 in Egyptian, which was slightly lower than were found in the previous studies (31) (20) (16). This prospective
283 study included all newly diagnosed children with ALL less than 15 years of age registered from October 2009 to
284 August 2014 at Radiation & Isotopes Center Khartoum versus 180 Egyptian ALL patients attending the Alksr
285 Aani, oncology center, Cairo. Besides epidemiological data, the objective of the current study was to look for
286 the age, sex, clinical features and, laboratory findings at presentation and compare it with reported literature.
287 Approximately 75% of ALL cases are in children and its after other nervous system and brain tumors (29). The
288 relationship between ALL and, age was report from the published sources found within this Literature Review
289 (30,32,33). As the ALL is the most common leukemia among children and the second most common cancer of
290 childhood after

291 14 Medical Research

292 Volume XXI Issue IV Version I(D D D D) F © 2021 Global Journals
293 Immunophenotyping of Acute Lymphoblastic Leukemia in Sudanese Children's Versus Egyptian Children's
294 using Flowcyometry

295 other nervous system and brain tumors, the current study was carried out in Sudanese and Egyptian children
296 with ALL (29) .A higher mortality rate was report among children with ALL, who were younger than two Yrs and
297 older than 10 Yrs (34) Another study found that 2-6 Yrs children with ALL survived more than those who were
298 less than 2 Yrs and older than 10 Yrs (Ref). ALL in pediatric is treat based on risk factors, which is defined by
299 laboratory and clinical features, therapeutic approach can be provided for patients who have a lower probability
300 of long-term survival (14,32), so that this study highlight and interpreted the frequency of ALL subclasses among
301 Sudanese and Egyptian population in correlation with their ages, as discussed in (Table ??) With regards to B
302 lineage, the prevalence of BALL in 9-12 yrs group was significantly higher in Sudanese compared to Egyptian
303 ($p<0.05$), whereas the frequency of Pre B subclass among those who were less than 8Yrs was higher in Egyptian
304 than in Sudanese ($p<0.05$) (See Table ??2). This finding is similar to a reported study by (Smibert 1996 and
305 co-workers, (34) 2-6 Yrs children with ALL were survival more than those who were less than 2 Yrs and older than
306 10 Yrs (34). This poor outcome in infants may be related to the common occurrence of other poor prognostic
307 features in this group of patients, such as higher leucocytic counts, higher incidence of hepatosplenomegaly and,
308 immunophenotype (13). Patients under the age of three years were found to have significantly lower intelligence
309 quotients than patients who received the same treatment at an older age and a group of healthy children matched
310 for age, sex and, parental occupation.

311 As the intensity of treatment required for favorable outcome varies substantially among subsets of children
312 with ALL, the participant's ages of the two ethnic groups were compared to identify the prognosis of the ALL in
313 each one. With regards to the ages of patients with T lineage ALL (thymocyte and post thymocyte), there was
314 no significant variation ($p>0.05$) between the two ethnic groups, Sudanese and Egyptian.(Table ??) summarized
315 gender differences in incidence rates of childhood Band T precursors ALL. Of B lineage ALL, males had higher
316 susceptibility to having ALL rather than females in both ethnic groups, still, in T lineage ALL, the frequency
317 was nearly the same in males and, females of both ethnic groups without significant variation (p -value > 0.05)
318 [See ??able 2]. Ching-Hon Pui and , coworkers were reported that boys had higher susceptibility to having T-cell
319 ALL than girls (20.9% v 10.7%) (35). This is in agreement with the current data, as males were more likely
320 to have T-cell ALL than females (66% v 34%, $P<.001$), in both subclasses of T-lineage ALL, thymocyte and
321 post-thymocyte. Acute lymphoblastic leukemia (ALL) develops at a rapidly, creating immature white blood cells
322 (WBCs) called lymphocytes. This type of hematological malignancy is cancer involving blood and bone marrow
323 (BM) (36).With regards to laboratory data, the Egyptian population showed severe anemia ($Hb < 8.8$ g/dl),
324 thrombocytopenia (68×10^3 /cm) and, ~%69 of them had blast cells. Interestingly, the B-ALL subclass had a
325 higher number of blast cells than other subclasses of ALL and, the thymocyte subclass had a higher percentage
326 of blast cells rather than the post thymocyte subclass (See Table3.5). This data was in agreement with a study
327 of (??4), as they found a high percentage of blast cells in cases with thymocyte & pre B ALL compared with
328 other subclasses of ALL. This may reflect that Egyptian patients with ALL have severe clinical symptoms, as
329 they might have a bleeding tendency because of low platelet and anemia as due to low Hb concentration.

330 Recurrent infection is not unexpected in those patients as they had a high number of blast cells in their
331 blood circulation. In contrast, Hb and platelets were found low among Sudanese cases, < 9 g/dl and 65×10^3
332 /cm, respectively which were the same as in the Egyptian population without significant variation ($p> 0.05$).
333 Blast cells were detect slightly higher among the Sudanese compared to the Egyptian, ~%78 without significant
334 variation ($p> 0.05$) [See Table ??].The presence of lymphadenopathy is usually found in cases with poor prognosis
335 and, patients who have a diagnostic problem (37). Thus it is important to distinguish between cases of ALL
336 with lymphadenitis from others. Clinical features such as lymph node (LN) enlargement and bleeding were taken
337 from each patient using a questionnaire. The presence of LN enlargement was detect significantly higher (p -value
338 <0.05) in Egyptian 85% ($N=153$) than in Sudanese 52% ($N=52$). Of Egyptian cases, its presence in B lineage
339 was found high pre B ALL phenotype 44% followed by pro B ALL 19.5% ($N=35$) and B ALL 7% ($N=14$) and its
340 presence in T lineage ALL was detected in all cases with post thymocyte and, its approximately three-quarters
341 of cases with thymocyte. Regarding lymphadenopathy among Sudanese, it was found that its presence and
342 absence were nearly the same, 52% and 48%, respectively. There was no significance variation in the presence

343 of lymph node enlargement between the B lineage ALL phenotypes in Sudanese. Interestingly, patients with
344 post thymocyte are possible to have lymphnode enlargement, as found that all Egyptian with post thymocyte
345 had lymphadenopathy and Sudanese patient with post thymocyte had lymphadenopathy except one case (See
346 Table3.6). Therefore, patients with thymocyte ALL phenotype might have a better outcomes in comparison
347 with post thymocyte ALL phenotype. The presence of bleeding was nearly equal in ethnic groups, 53% in
348 Sudanese and 54% in Egyptian. Regarding to bleeding in the study, population, the high frequency was found
349 mostly in the early phenotypes of B lineage ALL in Egyptian and, vice versa in Sudanese patients. Of T lineage
350 ALL, the presence of bleeding was common among Sudanese in both phenotype equally, thymocyte and post
351 thymocyte, but its presence in Egyptian was found mostly in thymocyte (See Table ??). In this study Sudanese
352 and Egyptian patients with ALL were classified into three categories based on their ages, (1-4 Yrs), (5-8 Yrs)
353 and the (9)(10)(11)(12). This Table describes two hundred eighty study participants, with ALL, of whom 36%
354 (n=100) were Sudanese and 64% (n=180) were Egyptian. The frequency of (1-4 yrs) age group was significantly
355 higher in the Egyptian than in the Sudanese ($p<0.05$). In contrast, the prevalence of (9-12 yrs) age group was
356 significantly higher in Sudanese compared to Egyptian ($p<0.05$). The Pre-BALL was significantly higher in
357 Egyptian than in Sudanese ($p<0.05$), particularly in those less than 8Yrs, whereas BALL was significantly higher
358 in Sudanese in comparison with Egyptian ($p<0.05$). With regards to the prevalence of T lineage ALL (thymocyte
359 and post thymocyte) and cases of B lineage ALL cases (Pro, Pre and, BALL), there was no significant variation
360 ($p>0.05$) between the two ethnic groups Sudanese and Egyptian. Their significance is represent in parenthesis.

361 **15 Medical**

362 **16 V. Conclusions**

363 This study concluded that:-Age ranging from one year to 12 years with a mean of 6.5 years. The male to female
364 ratio was 1:37 High age group in Egyptian ethnic group was 1-4 Yrs while in the Sudanese ethnic group the ,
365 higher age group was 9-12 Yrs. In this study, B lineage origin is most common type than T lineage origin in
366 two ethnic groups; In the T lineage had a better prognosis than B lineage. In this study, also thymocyte ALL
367 with cytoplasmic CD3 in the pediatric group below two years showed with a high total leucocytic count. Flow
368 cytometry has a distinctive role in the diagnosis and differentiation of ALL. using of certain flow cytometric
369 parameters can helps in minimization of cost without reduced accuracy. There is significant variations in ALL
370 sub classification between Sudanese and Egyptian Patients that may be due to genetic background.

371 **17 List of abbreviations**

372 AL

373 Figure Legends

374 1

.1 Declarations Ethical approval and consent to participant:

375 .1 Declarations Ethical approval and consent to participant:

376 Approval of This study was obtained from the hematology department of medical laboratory science (MLS),
377 Omdurman Islamic University and, the ministry of health issued by the local ethical committee Khartoum State,
378 Sudan. Written consent was taken from each member of the study.

379 .2 Consent for publication

380 Not applicable.

381 .3 Availability of data and materials

382 The datasets generated during and analyzed in this study are not publicly available due to Bahri hospital centers
383 ethical policy to protect participant confidentiality.

384 .4 Competing interest

385 The authors declare that they have no competing interests.

386 .5 Funding

387 .6 No funding was obtained for this study

388 .7 Authors contributions

389 AA and KO and AH contributed in literature search and manuscript writing. AA had the main idea of the study
390 and contributed in manuscript writing, KO contributed to clinic work, AH contributed in statistical analysis.
391 KO and AA supervised the study and critically reviewed the manuscript. All authors read and approved the
392 final draft of the manuscript.

393 [Bruske-Hohlfeld et al. ()] 'A cluster of childhood leukaemias near two neighbouring nuclear installations in
394 Northern Germany: prevalence of chromosomal aberrations in peripheral blood lymphocytes'. I Bruske-
395 Hohlfeld , H Scherb , M Bauchinger , E Schmid , H Fender , G Wolf . *Int J Radiat Biol* 2001. 77 (1) p.
396 .

397 [Ono et al. ()] 'A new strategy for treating patients with CD7+, CD4-, CD8-acute lymphoblastic leukaemia'. K
398 Ono , T Yoshida , K Tsuchiya , S Nakazato , F Nagumo , Y Shimamoto . *Eur J Haematol* 1997. 58 (2) p. .

399 [Seliger et al. ()] 'A rapid high-precision flow cytometry based technique for total white blood cell counting in
400 chickens'. C Seliger , B Schaefer , M Kohn , H Pendl , S Weigend , B Kaspers . *Vet Immunol Immunopathol*
401 2012. 145 (1-2) p. .

402 [Redaelli et al. ()] 'A systematic literature review of the clinical and epidemiological burden of acute lymphoblastic
403 leukaemia (ALL)'. A Redaelli , B L Laskin , J M Stephens , M F Botteman , C L Pashos . *Eur J Cancer
404 Care (Engl)* 2005. 14 (1) p. .

405 [Sen et al. ()] 'Acute lymphoblastic leukaemia (ALL) with infiltration of the thyroid: a cytological diagnosis'. R
406 Sen , S Gupta , A Batra , M Gill , V Gupta , N Marwah . *Endocr Pathol* 2012. 23 (4) p. .

407 [Pui ()] 'Acute lymphoblastic leukemia: introduction'. C H Pui . *Semin Hematol* 2009. 46 (1) p. .

408 [Yang et al. ()] 'Antileukaemia effect of rapamycin alone or in combination with daunorubicin on Ph+ acute
409 lymphoblastic leukaemia cell line'. X Yang , J Lin , Y Gong , H Ma , X Shuai , R Zhou . *Hematol Oncol* 2012.
410 30 (3) p. .

411 [Mushtaq et al. ()] 'Childhood acute lymphoblastic leukaemia: experience from a single tertiary care facility of
412 Pakistan'. N Mushtaq , Z Fadoo , A Naqvi . *J Pak Med Assoc* 2013. 63 (11) p. .

413 [Yasmeen and Ashraf ()] 'Childhood acute lymphoblastic leukaemia; epidemiology and clinicopathological fea-
414 tures'. N Yasmeen , S Ashraf . *J Pak Med Assoc* 2009. 59 (3) p. .

415 [Hashimabdulsalam et al. ()] 'Complementarity of evaluation of myeloperoxidase expression by flow cytometry
416 and immunohistochemistry on bone marrow trephine biopsy sections in acute myeloid leukaemia'. A
417 Hashimabdulsalam , E Nadal-Melsio , K N Naresh . *Cytometry B Clin Cytom* 2013.

418 [Griesinger et al. ()] 'Detection of HRX-FEL fusion transcripts in pre-pre-B-ALL with and without cytogenetic
419 demonstration of t (4;11)'. F Griesinger , H Elfers , W D Ludwig , M Falk , H Rieder , J Harbott . *Leukemia*
420 1994. 8 (4) p. .

421 [Kroll et al. ()] 'Evidence for under-diagnosis of childhood acute lymphoblastic leukaemia in poorer communities
422 within Great Britain'. M E Kroll , C A Stiller , S Richards , C Mitchell , L M Carpenter . *Br J Cancer* 2012.
423 106 (9) p. .

424 [Ahmed et al. ()] 'Five-year retrospective study of childhood leukaemia in the Sudan'. M A Ahmed , G A
425 Sulieman , A Omer . *East Afr Med J* 1977. 54 (1) p. .

17 LIST OF ABBREVIATIONS

426 [Iwamoto et al. ()] 'Flow cytometric analysis of de novo acute lymphoblastic leukemia in childhood: report from
427 the Japanese Pediatric Leukemia/Lymphoma Study Group'. S Iwamoto , T Deguchi , H Ohta , N Kiyokawa
428 , M Tsurusawa , T Yamada . *Int J Hematol* 2011. 94 (2) p. .

429 [Alves et al. ()] 'Flow cytometry immunophenotyping evaluation in acute lymphoblastic leukemia: correlation
430 to factors affecting clinic outcome'. G V Alves , A L Fernandes , J M Freire , Paiva Ade , S Vasconcelos , R
431 C Sales , VS . *J Clin Lab Anal* 2012. 26 (6) p. .

432 [Guy et al. ()] 'Flow cytometry thresholds of myeloperoxidase detection to discriminate between acute lym-
433 phoblastic or myeloblastic leukaemia'. J Guy , I Antony-Debre , E Benayoun , I Arnoux , C Fossat , Le
434 Garff-Tavernier , M . *Br J Haematol* 2013. 161 (4) p. .

435 [Jain et al. ()] 'Flow cytometry: a rapid and robust adjuvant technique for pathological diagnosis'. M Jain , A
436 Handoo , D R Choudhary , A Bhasin . *Turk J Haematol* 2012. 29 (1) p. .

437 [Kita et al. ()] 'Frequent gene expression of granulocyte colony-stimulating factor (G-CSF) receptor in CD7+
438 surface CD3-acute lymphoblastic leukaemia'. K Kita , K Nishii , K Ohishi , N Morita , N Takakura , K
439 Kawakami . *Leukemia* 1993. 7 (8) p. .

440 [Ferrando et al. ()] 'Gene expression signatures define novel oncogenic pathways in T cell acute lymphoblastic
441 leukemia'. A A Ferrando , D S Neuberg , J Staunton , M L Loh , C Huard , S C Raimondi . *Cancer Cell*
442 2002. 1 (1) p. .

443 [Nowicki et al. ()] 'In vitro substance P-dependent induction of bone marrow cells in common (CD10) acute
444 lymphoblastic leukaemia'. M Nowicki , D Ostalska-Nowicka , B Miskowiak . *Leuk Res* 2008. 32 (1) p. .

445 [Ahmed et al. ()] 'Leukaemia in the Democratic Republic of Sudan'. M A Ahmed , A A Kordofani , A Hidaytalla
446 , A Omer . *East Afr Med J* 1982. 59 (8) p. .

447 [Yoneda et al. ()] 'Lineage determination of CD7+ CD5-CD2-and CD7+ CD5+ CD2-lymphoblasts: studies on
448 phenotype, genotype, and gene expression of myeloperoxidase, CD3 epsilon, and CD3 delta'. N Yoneda , E
449 Tatsumi , K Teshigawara , S Nagata , T Nagano , Y Kishimoto . *Am J Hematol* 1994. 45 (4) p. .

450 [Alanio et al. ()] 'Low prevalence of resistance to azoles in Aspergillus fumigatus in a French cohort of patients
451 treated for haematological malignancies'. A Alanio , E Sitterle , M Liance , C Farrugia , F Foulet , F Botterel
452 . *J Antimicrob Chemother* 2011. 66 (2) p. .

453 [Karanth et al. ()] 'Mediastinal lymphadenopathy in a patient with previously treated T-cell acute lymphoblastic
454 leukaemia'. N Karanth , K P Prabhush , P N Karanth , T Shet , S D Banavali , P Parikh . *Med J Aust* 2008.
455 188 (2) p. .

456 [Hakulinen ()] 'Methodological problems in comparing incidence and prevalence of leukaemias and lymphomas:
457 ascertainment and age adjustment'. T Hakulinen . *Leukemia* 1999. 13 (1) p. . (Suppl)

458 [Carroll et al. ()] 'Pediatric acute lymphoblastic leukemia'. W L Carroll , D Bhojwani , D J Min , E Raetz , M
459 Relling , S Davies . *Hematology Am Soc Hematol Educ Program* 2003. p. .

460 [Seligmann et al. ()] 'Proceedings: B and T cell membrane markers in human leukaemias and lymphomata'. M
461 Seligmann , J C Brouet , J L Preud'homme . *Br J Cancer* 1975. 32 (2) p. .

462 [Settin et al. ()] 'Prognostic cytogenetic markers in childhood acute lymphoblastic leukemia'. A Settin , Al
463 Haggard , M , Al Dosoky , T , Al Baz , R Abdelrazik , N Fouada , M . *Indian J Pediatr* 2007. 74 (3) p.
464 .

465 [Settin et al. ()] 'Prognostic cytogenetic markers in childhood acute lymphoblastic leukemia: cases from Man-
466 soura Egypt'. A Settin , Al Haggard , M , Al Dosoky , T , Al Baz , R Abdelrazik , N Fouada , M . *Hematology*
467 2007. 12 (2) p. .

468 [Smibert et al. ()] 'Risk factors for intellectual and educational sequelae of cranial irradiation in childhood acute
469 lymphoblastic leukaemia'. E Smibert , V Anderson , T Godber , H Ekert . *Br J Cancer* 1996. 73 (6) p. .

470 [Schultz et al. ()] 'Risk-and responsebased classification of childhood B-precursor acute lymphoblastic leukemia:
471 a combined analysis of prognostic markers from the Pediatric Oncology Group (POG) and Children's Cancer
472 Group (CCG)'. K R Schultz , D J Pullen , H N Sather , J J Shuster , M Devidas , M J Borowitz . *Blood* 2007.
473 109 (3) p. .

474 [Mok et al. ()] 'RUNX1 point mutations potentially identify a subset of early immature T-cell acute lymphoblas-
475 tic leukaemia that may originate from differentiated T-cells'. M M Mok , L Du , C Q Wang , V Tergaonkar ,
476 T C Liu , Yin Kham , SK . *Gene* 2014. 545 (1) p. .

477 [Mecucci et al. ()] 't(4;11)(q21;p15) translocation involving NUP98 and RAP1GDS1 genes: characterization of a
478 new subset of T acute lymphoblastic leukaemia'. C Mecucci , La Starza , R Negrini , M Sabbioni , S Crescenzi
479 , B Leoni , P . *Br J Haematol* 2000. 109 (4) p. .

480 [Spiers et al. ()] 'Tcell chronic lymphocytic leukaemias and T-cell lymphoma-leukaemia: heterogeneity and
481 anomalous cell markers'. A S Spiers , D A Lawrence , M Levine , H Weitzman . *Scand J Haematol* 1986. 37
482 (5) p. .

483 [Van Vlierberghe and Ferrando ()] 'The molecular basis of T cell acute lymphoblastic leukemia'. P Van Vlier-
484 berghe , A Ferrando . *J Clin Invest* 2012. 122 (10) p. .

485 [Flavell et al. ()] 'Therapy of human T-cell acute lymphoblastic leukaemia with a combination of anti-CD7 and
486 anti-CD38-SAPORIN immunotoxins is significantly better than therapy with each individual immunotoxin'.
487 D J Flavell , D A Boehm , A Noss , S L Warnes , S U Flavell . *Br J Cancer* 2001. 84 (4) p. .

488 [Victor Hoffbrand et al. ()] A Victor Hoffbrand , Dc , G D Edward , Anthony R Tuddenham , Green .
489 *Postgraduate Haematology*, (London) 2010. Wiley-Blackwell. (6th ed)